



Maharashtra Education Society's
Abasaheb Garware College
(Autonomous)

(Affiliated to Savitribai Phule Pune University)

Three Year B.Sc. Degree Program in Microbiology
(Faculty of Science and Technology)

Syllabi under Autonomy
F.Y.B.Sc. (Microbiology)

Choice Based Credit System Syllabus
To be implemented from Academic Year 2022-2023

Title of the course: B.Sc. (Microbiology)

Preamble:

Microbiology is a broad discipline of biology which encompasses five groups of microorganisms i.e., bacteria, protozoa, algae, fungi, viruses. It studies their interaction with their environments as well as how these organisms are harnessed in human endeavour and their impact on society. The study has its extensions in various other conventional and advanced fields of biology by employing microbes as study models. Since inception of microbiology as a branch of science, it has remained an ever-expanding field of active research, broadly categorized as pure and applied science. Microorganisms were discovered over three fifty years ago and it is thought that a huge diversity yet remains to be explored.

Knowledge of different aspects of Microbiology has become crucial and indispensable to the society. Study of microbes has become an integral part of education and human progress. There is a continuous demand for microbiologists as work force – education, industry and research. Career opportunities for the graduate students are available in industry and research equally.

Program outcome:

- To enrich students' knowledge and train them in the pure microbial sciences
- To introduce the concepts of application and research in Microbiology
- To inculcate sense of scientific responsibilities and social and environment awareness
- To help students build-up a progressive and successful career

Eligibility for Admission:

First Year B.Sc.:

Higher Secondary School Certificate (10+2) or its equivalent Examination with English and Biology; and two of the science subjects such as Physics, Chemistry, Mathematics, Geography, Geology, etc.

Admissions will be given as per the selection procedure / policies adopted by the college keeping in accordance with conditions laid down by the Savitribai Phule Pune University.

Reservation and relaxation will be as per the Government rules.

Structure of the Course: (Microbiology)

Year	Semester	Course Type	Course Code	Course Title	Remark	Credit	No. of Lectures /Practical to be conducted
1	I	Compulsory	USMR- 111	Introduction to Microbial World	Theory	2	36
		Compulsory	USMR- 112	Basic Techniques in Microbiology	Theory	2	36
		Compulsory	USMRP- 113	Practical Course based on theory paper I (USMR-111) and Paper II (USMR-112)	Practical	1.5	14
	II	Compulsory	USMR- 121	Bacterial Cell and Biochemistry	Theory	2	36
		Compulsory	USMR- 122	Microbial cultivation and growth	Theory	2	36
		Compulsory	USMRP- 123	Practical Course based on theory paper I (USMR-121) and Paper II (USMR-122)	Practical	1.5	14
2	III	Compulsory	USMR-231	Medical Microbiology and Immunology	Theory	2	36
		Compulsory	USMR-232	Bacterial physiology and Fermentation Technology	Theory	2	36
		Compulsory	USMRP-233	Practical based on USMR-231and USMR232	Practical	2	12
		Compulsory	USLG-231	Language	Theory	2	36
		Compulsory	AACC-231	Environmental Science	Theory	2	36
	IV	Compulsory	USMR-241	Bacterial Genetics	Theory	2	36
		Compulsory	USMR-242	Air, Water and Soil Microbiology	Theory	2	36
		Compulsory	USMRP-243	Practical basedon USMR-241and USMR-242	Practical	2	12
		Compulsory	USLG-241	Language	Theory	2	36
		Compulsory	AACC-241	Environmental Science	Theory	2	36
3	V	DSEC-Compulsory	USMR- 351	Medical Microbiology- I	Theory	2	36
		DSEC-Compulsory	USMR- 352	Immunology- I	Theory	2	36
		DSEC-Compulsory	USMR- 353	Enzymology	Theory	2	36
		DSEC-Compulsory	USMR- 354	Genetics	Theory	2	36
		DSEC-Compulsory	USMR- 355	Fermentation technology- I	Theory	2	36
		DSEC-Compulsory	USMR- 356	Agricultural Microbiology	Theory	2	36
		DSEC-Compulsory	USMRP- 357	Practical Course I	Practical	2	12

		DSEC-Compulsory	USMRP- 358	Practical Course II	Practical	2	12
		DSEC-Compulsory	USMRP- 359	Practical Course III	Practical	2	12
		SEC-Compulsory	USMRSEC-3510	Marine microbiology	Theory/practical	2	36
		SEC-Compulsory	USMRSEC-3511	Dairy Microbiology	Theory/practical	2	36
	VI	DSEC-Compulsory	USMR- 361	Medical Microbiology- II	Theory	2	36
		DSEC-Compulsory	USMR- 362	Immunology- II	Theory	2	36
		DSEC-Compulsory	USMR- 363	Metabolism	Theory	2	36
		DSEC-Compulsory	USMR-364	Molecular Biology	Theory	2	36
		DSEC-Compulsory	USMR- 365	Fermentation technology II	Theory	2	36
		DSEC-Compulsory	USMR- 366	Food Microbiology	Theory	2	36
		DSEC-Compulsory	USMRP- 367	Practical Course I	Practical	2	12
		DSEC-Compulsory	USMRP- 368	Practical Course II	Practical	2	12
		DSEC-Compulsory	USMRP- 369	Practical Course III	Practical	2	12
		SEC-Compulsory	USMRSEC-3610	Waste management	Theory/practical	2	36
		SEC-Compulsory	USMRSEC-3611	Nanobiotechnology	Theory/practical	2	36

DSEC: Discipline Specific Elective Course, SEC: Skill Enhanced Course

SEMESTER-I

Course code and title: USMR-111 Introduction to Microbial World

Lectures: (Credits- 02)

Course Outcomes:

1. Students will understand the contributions of different scientists in the fields of Microbial science.
2. Students will have knowledge about different types of microbes.
3. Students will have knowledge about the established and emerging fields of science with respect to Microbiology.

Credit	Topic	No. of Lectures (36)
Credit I	1. History of Microbiology in developing as a Modern fundamental Science.	
	❖ Discovery Era: Discovery of Microorganisms (Contributions of Anton von Leeuwenhoek and Robert Hooke).	2
	❖ Transition Era: Conflict of Abiogenesis v/s biogenesis theory	3
	❖ Golden Era of Microbiology:	4
	➤ Birth of Bacteriology, Medical Microbiology and Virology due to Contributions of following scientists: <ul style="list-style-type: none"> ○ Louis Pasteur (Fermentation, Pasteurization) ○ Contribution of Robert Koch (Germ theory of disease, Tuberculosis and Cholera, concept of Pure culture) ○ Ferdinand Cohn (Endospore discovery). ○ Discovery of viruses, River's Postulates. ○ Breakthroughs in medical field: Contribution of Joseph Lister (antiseptic surgery), Paul Ehrlich (Chemotherapy), Alexander Fleming (Penicillin), Discovery of Streptomycin by Walksman. 	3
	➤ Establishment of Immunology: Vaccination, First Laboratory Vaccine, Story of Rabies vaccine, contribution of Elie Metchnikoff (Phagocytosis).	3
	➤ Development of Soil Microbiology: Contribution of Martinus W. Beijerinck, Sergei N. Winogradsky.	1
	❖ Modern Era of Microbiology (Contribution of Carl Woese)	1
	2. Public Health and Microbiology: COVID -19 Pandemic	1
	<i>(Project Based Learning: Assignments should be given to student)</i>	

Credit II	2. Systems of Classification: Comparative account of Prokaryotes and Eukaryotes, Three domain classification and Five kingdom classification.	2
	3. Types of Microorganism and their differentiating characters (with emphasis on distribution and occurrence, morphology, classification, mode of reproduction and medical /economic/ environmental importance)	
	a. Bacteria (Eubacteria, Archaeobacteria, Actinomycetes)	
	b. Protozoa	6
	c. Fungi	
	d. Algae	
	e. Viruses (Classification of Viruses: ICTV nomenclature)	
	f. Viroids and Prions	
	3. Scope of studying microorganisms:	
	a. Bacterial Taxonomy - Introduction to Bergey's Manual of Systematics of Archae & Bacteria (BMSAB) and Bergey's International Society for Microbial Taxonomy (BISMIS).	2
b. Medical Microbiology	1	
c. Environmental Microbiology	1	
d. Food and Dairy Microbiology	1	
e. Agriculture Microbiology	1	
f. Industrial Microbiology	1	
g. Immunology	1	
h. Space Microbiology	1	
i. Nano Science	1	

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SEMESTER-I**Course code and title: USMR-112 Basic Techniques in Microbiology****Lectures: (Credits- 02)**

Course Outcomes:

1. Students will be able to understand the needs and basics of techniques used in observing microbes.
2. Students will be aware of applications of basic techniques.
3. Students will learn sterilization and disinfection principles and procedures.

Credit	Topic	No. of Lectures (36)
Credit I	1. Bacterial size, measurement and relevant techniques	1
	2. Microscopy	
	a. Bright field microscopy and Dark field Microscopy	4
	<ul style="list-style-type: none"> ● Electromagnetic spectrum of light, concepts of magnification, numerical aperture and resolving power. ray diagram ● Parts of microscope -condensers, eyepieces and objectives ● Concept of aberrations in lenses - spherical, chromatic 	
	b. Principle, working and ray diagram of	1
<ul style="list-style-type: none"> ● Phase contrast microscope ● Fluorescence Microscopy ● Electron Microscopy – TEM, SEM 	1	
		2

	<p>3. Staining Techniques: Concept and Need of staining</p> <p>a. Stain; Types (Basic and Acidic), Properties and role of Fixatives, Mordants, Decolourisers and Accentuators</p> <p>b. Monochrome staining and Negative (Relief) staining</p> <p>c. Differential staining - Gram staining and Acid-fast staining</p> <p>d. Special staining- Capsule (Maneval's method, Hiss method), Cell wall (Chance's method, Ringers's Method), Endospore (Dorner's method, Schaeffer -Fulton's method), Flagella (Leifson's method, Loeffler's method), metachromatic granules (Alberts Method)</p>	<p>1</p> <p>1</p> <p>1</p> <p>2</p> <p>4</p>
Credit II	<p>4. Sterilization and Disinfection</p> <p>a. Sterilization</p> <ul style="list-style-type: none"> ● Physical Agents - Heat, Radiation, Filtration ● Checking of efficiency of sterilization (Dry and Moist) – Biological and Chemical Indicators <p>b. Disinfection:</p> <ul style="list-style-type: none"> ● Chemical agents and their mode of action - Aldehydes, Halogens, Quaternary ammonium compounds, Phenol and phenolic compounds, ● Heavy metals, Alcohol, Dyes, Detergents and Ethylene oxide. ● Characteristics of an ideal disinfectant ● Checking of efficiency of disinfectant - Phenol Coefficient (Rideal-Walker method) ● Sanitization 	<p>3</p> <p>2</p> <p>4</p> <p>4</p> <p>1</p> <p>1</p> <p>1</p>

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SEMESTER-I**Course code and title: USMRP-113 Practical Course based on USMR-111 and USMR-112****Lectures: (Credits- 1.5)**

Course Outcomes:

1. Students will understand safety measures in microbiology laboratory.
2. Students will be Introduced to microbiology laboratory and common microbiology laboratory instruments.
3. Students will get acquainted with skills of aseptic culture technique
4. Students will observe microorganisms from natural samples.
5. Students will be able to prepare glassware and media for cultivation of microbes.

Expt. No.	Practical	No. of Practicals (14)
1	<p>a. Safety measures and Good Laboratory Practices in microbiology laboratory.</p> <p>b. Preparation of SOP and demonstration of operations of common microbiology laboratory instruments: Incubator, Hot air oven, Autoclave, Colorimeter, Laminar air flow hood, Clinical centrifuge.</p>	2
2	<p>a. Construction (mechanical and optical), working and care of bright field microscope.</p> <p>b. Permanent slide observation: Algae, Fungi and Protozoa</p> <p>c. Wet mount slide preparation of natural samples and observation for: Bacteria, Algae, Fungi and Protozoa.</p>	3
3	<p>a. Introduction and use of common laboratory glass wares: Test tubes, culture tubes, suspension tubes, screw capped tubes, Petri plates, pipettes (Mohr and serological) micropipettes, Pasteur pipettes, Erlenmeyer flask, volumetric flask, glass spreader, Durham's tube, Cragie's tube and inoculating needles (wire loop, stab needles).</p> <p>b. Learning basic techniques in Microbiology: Wrapping of glassware, cotton plugging, cleaning and washing of glassware, biological waste disposal.</p> <p>c. Aseptic culture Technique</p>	2

4	Basic staining techniques: a. Monochrome staining b. Negative staining c. Gram staining d. Acid-fast staining (ZNCF Method)	4
5	Observation of motility in bacteria using: Hanging drop method and swarming growth method.	2
6	Checking of efficacy of chemical disinfectant: Phenol Coefficient by Rideal-Walker method. (Demonstration)	1

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SEMESTER-II

Course code and title: USMR-121 Bacterial Cell and Biochemistry

Lectures: (Credits- 02)

Course Outcomes:

1. Students will be able to understand bacterial cell structure and functions.
2. Students will have knowledge about different types of biomolecules and their functions.
3. Students will know about communication between bacteria.

Credits	Topic	No. of Lectures (36)
Credit I	1. Ultrastructure of bacterial cell:	2
	a. Cell wall (Gram positive, Gram negative)	1
	b. Concept of Mycoplasma, Spheroplast, protoplast, L-form	2
	c. Cell membrane	1
	d. Endospore (spore formation and stages of sporulation)	1
	e. Capsule	1
	f. Flagella	2
	g. Fimbriae and Pili	2
	h. Ribosomes	1
	i. Chromosomal & extra-chromosomal material	1
	j. Cell inclusions (Gas vesicles, carboxysomes, PHB granules, metachromatic granules, glycogen bodies, starch granules, magnetosomes, sulfur granules, chlorosomes)	2
	2. Communication in bacteria through Quorum sensing	5
	1	

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SEMESTER-II**Course code and title: USMR-122: Microbial cultivation and growth****Lectures: (Credits- 02)**

Course Outcomes:

1. Students will have knowledge about microbial nutritional classification and microbial cultivation media.
2. Students will learn about the technique of isolation and enumeration of microbes.
3. Students will understand the bacterial growth kinetics and measurements.

Credit	Topic	No. of Lectures (36)
Credit I	1. Cultivation of Microorganisms:	
	a. Chemical composition of typical bacterial cell, Nutrition and Nutritional requirements, classification based on nutrition.	4
	b. Design and preparation of culture media: Common ingredients of media, types of culture media (Complex, synthetic, semi-synthetic, etc.)	3
	c. Concept of Enrichment, Pure Culture, Isolation of culture by streak plate, pour plate, spread plate.	3
	d. Methods for cultivating photosynthetic, extremophilic and chemo-lithotrophic bacteria, anaerobic bacteria, algae, fungi, actinomycetes and viruses.	4
	e. Maintenance of bacterial and fungal cultures using different techniques.	3
	f. Culture collection centres and their role.	1

Credit II	2. Bacterial growth:	
	a. Kinetics of bacterial growth	3
	Binary division, budding, fragmentation, Exponential growth model, calculation of generation time and specific growth rate	2
	b. Growth curve- phases, Diauxic growth and Synchronous growth	4
	c. Batch culture and Continuous culture, Concept of fermentation, yield- biomass and product	
	d. Measurement of bacterial growth- Microscopic methods -Direct microscopic count, counting cells using Neubauer chamber	2
	e. Plate counts (Total viable count)	2
	f. Turbidometric methods and biomass (Dry mass)	2
g. Factors affecting bacterial growth [pH, Temperature, Solute Concentration (Salt and Sugar)] and Heavy metals.	3	

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SEMESTER-II**Course code and title: USMRP-123 Practical Course based on USMR-121 and USMR-122****Lectures: (Credits- 1.5)**

Course Outcomes:

1. Students will be able to prepare microbial cultivation media and acquire skills of isolations techniques.
2. Students will be able to stain bacteria by special techniques.
3. Students will be aware of preservation and maintenance of microbes in laboratory conditions.

Expt No.	Topics	No. of practical (14)
1	i. Preparation of simple laboratory nutrient media (Nutrient agar/broth, MacConkey's agar). ii. Checking sterilization efficiency of autoclave using a biological indicator (<i>B. stearothermophilus</i>) iii. Preparation of Winogradsky's column and observation of different types of microorganisms using bright field microscope	1 1 1
2	Special staining techniques: 1. Cell wall staining (Chance's method) 2. Capsule staining (Manevals method) 3. Endospore staining (Schaeffer -Fulton's method) 4. Metachromatic granules staining (Albert's Method)	1 1 1 1
3	Preservation of cultures on: Slants, glycerol stocks Demonstration of lyophilization and lyophilized cultures.	1
4	Isolation of bacteria: Streak plate technique (Colony characteristics)	1

5	Enumeration of bacteria from fermented food / soil / water by: i. Spread plate method ii. Pour plate method	2
6	To study the effect of different parameters on growth of <i>Bacteria</i>: i. pH, temperature, sodium chloride concentration, Oxygen conditions. ii. Study of oligodynamic action of heavy metal	2
7	Growth curve of bacteria	1

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